Disease Resistance and Nutritional Properties of Tuber-Bearing Native Potato Species and Old Spanish Cultivars

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ABSTRACT

Certain potato cultivars such as native potato species (NPS) from the Andes are known to have resistances to different pests and diseases. Some accessions are also interesting from a nutritional and culinary perspective. A collection of 35 NPS and 11 old Spanish accessions were analysed for *Streptomyces scabies*, *Rhizoctonia solani* and *Globodera rostochiensis*, as well as dry matter, reducing sugars, minerals, glycoalkaloid concentrations, and total antioxidant capacity. A wide variability was found between and also within the species. Most accessions showed favourable characteristics, while high concentrations values of glycoalkaloids were observed in certain accessions. The results suggest that some NPS and old Spanish accessions have a great potential for exploitation in potato breeding programmes as a source of resistances and nutritional variability.

Keywords: Antioxidant capacity, Breeding, Glycoalkaloids, Pathogens.

INTRODUCTION

Potato is a basic food crop in many countries that produces a higher amount of energy and protein per surface area than any other food crop, including cereals. In addition, potatoes are a valuable source of carbohydrates, proteins, minerals (particularly calcium and potassium) and vitamins (Fernie and Willmitzer, 2001). Potato has probably more related wild species than any other crop, since the genus Solanum comprises around 2,000 species (Ruiz de Galarreta and Ríos, 2008). Among them are 189 tuber-bearing species, including one cultivated species (Spooner and Salas, 2006). It was first cultivated 6,000 to 10,000 years ago around the Titicaca Lake. In successive generations, the Andean communities obtained hundreds of cultivars, extending the potato culture to the whole Andean region and occupying the high regions of South America (Spooner and Hetterscheid, 2006). The current potato diversity is wide and more than 5,000 landraces of potatoes have been identified by the International Potato Centre (Huamán, 1986), with different shapes, colors, and chemical compositions (De Haan, 2006).

Despite some introgressions from other *Solanum* species, modern potato cultivars belong basically to the species *Solanum tuberosum* ssp. *tuberosum*. However, in South-America other tuber-bearing species are cultivated, which are called Native Potato Species (NPS). They include figure *S. juzepczukii* (3x), *S. chaucha* (3x), *S. stenotonum* (2x), *S. ajanhuiri* (2x) with the groups Yari and Ajawiri, *S. goniocalyx* (2x), *S. curtilobum* (5x), *S. phureja* (2x) and *S. tuberosum ssp. andigena* (4x). The native germplasm presents remarkable genetic diversity (Spooner *et al.*, 2005), including resistances to pest and diseases (Gabriel *et*

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al., 2011) and high nutritional variability (Burgos *et al.*, 2007).

In spite of its common perception as a carbohydrate source, potatoes, particularly NPS, are also a good source of relative high quality proteins (Dijkstra et al., 2003). Some of them are also great sources of vitamins, dietary fibre, and minerals (Lu et al., 2001). Potato tubers also contain a variety of phytonutrients, including carotenoids and phenolic compounds. The content of bioactive compounds is affected by several environmental factors, but, in most cases, the genotype has a determining effect (Brown et al., 1993). Although potato antioxidant capacity is low when compared to other vegetables (Stratil et al., 2006), Solanum cultivated species exhibit a wide variation in total antioxidant capacity values, and higher values are usually found in genotypes with deep colored skin and flesh. Purple or red potatoes are especially rich in secondary metabolites like anthocyanins. In addition, yellow and orange fleshed potatoes are particularly rich in certain xantophyll carotenoids, including type lutein. violaxanthin and zeaxanthin (Brown, 2005).

However, most accessions of NPS present also some undesirable agronomic, nutritional, or commercial characteristics. The absence of dormancy and the presence of excessively long stolons are considered as adverse factors that appear in certain native potatoes. Extremely high total glycoalkaloid constitute concentrations also an unfavourable characteristic of certain NPS accessions. Glycoalkaloids represent a toxic group of secondary plant compounds found throughout the foliage and tubers of Solanaceae (Friedman, 2004). They are thought to function as the chemical defence of the plant against potential pathogens (Friedman and McDonald, 1997). The two major glycoalkaloids in potatoes are α solanine and α -chaconine, but the ratio of α solanine to α -chaconine varies widely among tissues, genotypes, and growing conditions (Sarquis et al., 2000). These steroidal compounds are reported to exhibit both beneficial and adverse effects (Friedman *et al.*, 2005). In small amounts, these compounds contribute to potato flavour, but potatoes containing glycoalkaloids in excess are unsafe for human consumption (Van Gelder, 1988).

Following a previous paper on characterization of NPS (Ritter *et al.*, 2008), the objective of the present study was to analyze additional traits of NPS and certain old Spanish accessions including resistances to *Streptomyces scabies*, *Rhizoctonia solani*, and *Globodera rostochiensis*, dry matter, reducing sugars and minerals, as well as the concentration of glycoalkaloids and total antioxidant capacity.

MATERIALS AND METHODS

Plant Material

A total of 46 accessions representing different NPS and old Spanish potatoes belonging to Solanum tuberosum ssp. tuberosum were analyzed. Native potato accessions were acquired from International Potato Center (Lima, Peru). These were selected with the aim of maximizing the geographic coverage and genetic diversity. Spanish accessions were selected from the germplasm collection held at Neiker-Tecnalia. The commercial cultivar Desirée belonging to the species S. tuberosum was selected as susceptible controls in the evaluation of resistance and the cv. Agria as quality check. Details about plant material are shown in Table 1.

Evaluation of Resistances against Pathogens

Tuber resistance against *Streptomyces* scabies was determined by growing five potato plants of each accession in infested soil with 40 mL of the spore suspension of an isolate from Neiker Institute (Spain) during 10 weeks according to Bouchek-Mechiche *et al.* (1998). The resistance against *Rhizoctonia solani* was analyzed

| Accession | | | | | |
|--------------------|-----------------------------------|----------------------------|-----------------|------------|------------------------|
| code | Name | Species ^a | Origin | Skin color | Flesh color |
| NKD-138 | Laram Ajawiri | S. ajawiri | Bolivia | Yellow | Yellow |
| NKD-139 | Jancko Ajawiri | S. ajawiri | Bolivia | Yellow | White |
| NKD-126 | Ojo de Buey | S. andigena | Peru | Brown | White |
| NKD-128 | Huagalina | S. andigena | Peru | Brown | Yellow |
| NKD-130 | Muro Shocco | S. andigena | Peru | Purple | Yellow-Purple |
| NKD-134 | Pulu | S. andigena | Bolivia | Purple | White-Purple |
| NKD-135 | Socco Huaccoto | S. andigena | Peru | Purple | Yellow |
| NKD-137 | Sipancachi | S. andigena | Bolivia | Yellow | Yellow |
| NKD-141 | Unknown-1 | S. andigena | Peru | Yellow | Yellow |
| NKD-143 | Wila Huaka Lajra | S. andigena | Bolivia | Red | Yellow |
| NKD-145 | Puca quitish | S. andigena | Peru | Purple | Purple |
| NKD-156 | Holandesa | S. andigena | Colombia | Red | Yellow |
| NKD-150 NKD-157 | Unknown-2 | S. andigena S. andigena | Colombia | White | White |
| | Camusa | | Venezuela | | |
| NKD-159 NKD 160 | Camusa Chimbina | S. andigena | Peru | Purple | White-Purple |
| NKD-160 | | S. andigena | Peru Peru | Purple | Yellow-Purple White |
| NKD-161 NKD-151 | Negrita Chiar Surimana o Phiñu | S. andigena S. chaucha | Peru Bolivia | Purple | White |
| | Color Unckuna | | | Purple | |
| NKD-163 | | S. chaucha | Peru | Red | Yellow-Red |
| NKD-131 | Puca Huayro | S. chaucha | Peru | Red | Yellow-Red |
| NKD-153 | Unknown-3 | S. goniocalyx | Peru | Brown | Yellow |
| NKD-155 | Kashpadana Amarilla | S. goniocalyx | Peru | Red | Yellow |
| NKD-133 | Chimi Lucki | S. juzepczukii | Bolivia | White | White |
| NKD-144 | Laram Canchali | S. juzepczukii | Peru | Purple | White |
| NKD-132 | Chaucha | S. phureja | Peru | Purple | Yellow |
| NKD-150 | Rosca | S. phureja | Colombia | Yellow | Yellow |
| NKD-127 | Calhua Rosada | S. stenotonum | Peru | Brown | Yellow |
| NKD-129 | Señora Warni | S. stenotonum | Peru | Purple | Yellow |
| NKD-140 | Morar Nayra Mari | S. stenotonum | Peru | Red | Yellow-Purple |
| NKD-142 | Yana Sucre | S. stenotonum | Peru | Purple | White |
| NKD-148 | Cceccorani | S. stenotonum | Peru | Brown | Yellow-Purple |
| NKD-149 | Yana Ppoccoya | S. stenotonum | Peru | Red | White |
| NKD-152 | Morada Turuna | S. stenotonum | Peru | Red | Yellow-Red |
| NKD-158 | Poluya | S. stenotonum | Peru | Purple | Yellow |
| NKD-164 | Amarilla | S. stenotonum | Peru | Yellow | Yellow |
| NKD-154 | Ucho Chaquitay | SxG ^b | Colombia | Purple | Yellow-Purple |
| NK-272 | Cazona | S. tuberosum | Spain | Yellow | Yellow |
| NK-273 | Fina de Carballo | S. tuberosum | Spain | Yellow | Yellow |
| NK-520 | Fina de Gredos | S. tuberosum | Spain | Brown | White |
| NK-136 | Kasta | S. tuberosum | Spain | Purple | Purple |
| NK-129 | Jesus | S. tuberosum | Spain | Purple | Purple |
| NK-338 | Morada | S. tuberosum | Spain | Purple | Purple |
| NK-222 | Roja Riñon | S. tuberosum | Spain | Red | Yellow |
| NK-011 | Alegria Oro | S. tuberosum | Spain | Yellow | Yellow |
| NK-292 | Ibicenca | S. tuberosum | Spain | Red | Yellow |
| NK-201 | Pedro Muñoz | S. tuberosum | Spain | Yellow | White |
| NK-515 | Tramontana | S. tuberosum | Spain | Yellow | Yellow |
| NK-172 | Agria ^c | S. tuberosum | Germany | Yellow | Yellow |
| NK-069 | Desiree ^c | S. tuberosum | Netherlands | Red | Yellow |

Table 1. Native Potato Species (NPS) and old Spanish accessions evaluated.

^a S. tuberosum=S. tuberosum ssp. tuberosum, S. andigena=S. tuberosum ssp. andigena

^b Hybrid between S. tuberosum ssp. andigena and S. goniocalyx., ^c Varieties used as controls

using five tubers of each accession inoculated in pot with under greenhouse conditions during 10 weeks at 20 °C with 16 h light period (Little *et al.*, 1988). The evaluation of both resistances was based on a damage scale established by James (1971) that considers the affected tuber area. The presence of black or brown sclerotia on the tuber surface was evaluated. Partial resistance was considered if less then 10% or less then 5% of the tuber surface was covered by sclerotia from *S. scabies* and *R. solani*, respectively.

For detecting resistance against the nematode Globodera rostochiensis, virulence group Ro1, a bioassay in the greenhouse using five tubers as replicates was performed as a pot test with infected soil with 30 cysts per pot leading to an infestation density of 10 to 20 juveniles per gram of soil following the methodology described by Ruiz de Galarreta et al. (1998). Pots were watered lightly and evenly and plants were grown at 20 °C for 10 weeks. Susceptible accessions were identified visually by observing females on the roots according to Rousselle-Bourgeois and Mugniéry (1995). An accession was defined as resistant, if less than 5 cysts appeared in each of the replicates, and as partially resistant, if multiplication rates were less than 10% compared to the susceptible control in each pot.

Determination of Dry Matter, Reducing Sugars, and Minerals

Estimation of dry matter (DM) after drying was performed according to Gould (1999) and calculated as the ratio between dry and fresh mass. Reducing sugars were quantified using standardized а spectrophotometrical procedure with DNS (3, 5-Dinitrosalicylic acid) as reactive (Sumner, 1921). Total reducing sugar concentration (RS) was calculated following the relation between absorbance and sugar content described by Barredo and Ritter (1992). Samples were analyzed for P, Ca, K, Mg, Na, Zn, and Fe using ICP-OES spectrometry (AOAC, 2012). Mineral composition was evaluated by acid digestion at 140 °C in 70% HNO₃/HClO₄ and ICP-OES using an ARL 3580B ICP (ARL, Switzerland). Least significant differences were estimated. All analyses were performed with the SAS package.

Antioxidant Capacity and Glycoalkaloids

The total antioxidant capacity (TAC) of potato tubers was determined by ORAC (Oxygen Radical Absorbance Capacity) standardized assay (Cao et al., 1993). In the ORAC method, a sample is added to the peroxyl radical generator AAPH (2,2'-Azobis-(2-Amidinopropane)-dihydrochloride) and inhibition of the free radical action is measured using the fluorescent compound, Bphycoerythrin or R-phycoerythrin (Cao et al., 1995). Glycoalkaloids were analyzed by extraction with acetic acid and determined by HPLC. Lyophilized, completely desprouted, and ground samples were extracted with the extraction solution (water/acetic acid/ sodium hydrogen sulphite 95+5+0.5, v/v/w) for 15 min (Hellenäs et al., 1995). Glycoalkaloids were eluted with the HPLC mobile phase (acetonitrile/0.022 M potassium phosphate buffer, pH 7.6, 55:45 v/v). The HPLC separation was carried out by a HyperClone 5 µm ODS phase (C18; Phenomenex) in a 150×4.6-mm column. Quantification was made by UV-detection at 202 nm using a flow rate of 1.5 mL/min (Knauer Instruments, HPLC pump Smartline 1000) and comparison with areas of the external standards, α -solanine α -chaconine. Total glycoalkaloid and concentration (TGA) results were calculated and expressed as mg TGA/kg fresh weigh (FW), by summing up the contents of α solanine and α -chaconine (Haase, 2010).

RESULTS

The results of resistance screenings against *S. scabies*, *R. solani* and *G. rostochensis* are shown in Table 2. Only 12 accessions were

| NKD-138 10 5 S NKD-139 25 5 S NKD-126 0 10 S NKD-128 10 1 S NKD-130 10 10 S NKD-130 10 10 S NKD-134 0 5 S NKD-135 50 10 S NKD-137 50 15 S NKD-141 10 10 S NKD-143 10 10 S NKD-143 10 10 S NKD-145 10 1 S NKD-156 25 10 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-151 25 10 S NKD-163 25 10 S NKD-153 10 10 S | 22.0±1.3* 23.9±1.1 21.6±1.0 21.8±1.3 23.2±0.9 24.9±0.9 19.7±1.3 23.8±1.0 23.2±0.8 26.3±0.6 21.2±1.3 22.7±1.3 29.3±1.1 | $1.10\pm0.12*$ 1.05 ± 0.10 1.01 ± 0.18 0.66 ± 0.15 0.87 ± 0.15 0.49 ± 0.10 0.59 ± 0.13 0.15 ± 0.12 0.17 ± 0.13 0.37 ± 0.11 1.10 ± 0.16 |
|--|---|--|
| NKD-126 0 10 S NKD-128 10 1 S NKD-130 10 10 S NKD-134 0 5 S NKD-135 50 10 S NKD-137 50 15 S NKD-141 10 10 S NKD-143 10 10 S NKD-143 10 10 S NKD-145 10 1 S NKD-156 25 10 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 S NKD-151 25 10 S NKD-163 25 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR | 21.6±1.0 21.8±1.3 23.2±0.9 24.9±0.9 19.7±1.3 23.8±1.0 23.2±0.8 26.3±0.6 21.2±1.3 22.7±1.3 29.3±1.1 | 1.01±0.18 0.66±0.15 0.87±0.15 0.49±0.10 0.59±0.13 0.15±0.12 0.17±0.13 0.37±0.11 |
| NKD-128 10 1 S NKD-130 10 10 S NKD-134 0 5 S NKD-135 50 10 S NKD-137 50 15 S NKD-141 10 10 S NKD-143 10 10 S NKD-145 10 1 S NKD-156 25 10 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 PR NKD-151 25 10 S NKD-163 25 10 S NKD-131 25 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 21.8±1.3 23.2±0.9 24.9±0.9 19.7±1.3 23.8±1.0 23.2±0.8 26.3±0.6 21.2±1.3 22.7±1.3 29.3±1.1 | 0.66±0.15 0.87±0.15 0.49±0.10 0.59±0.13 0.15±0.12 0.17±0.13 0.37±0.11 |
| NKD-128 10 1 S NKD-130 10 10 S NKD-134 0 5 S NKD-135 50 10 S NKD-137 50 15 S NKD-141 10 10 S NKD-143 10 10 S NKD-145 10 1 S NKD-156 25 10 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 PR NKD-151 25 10 S NKD-163 25 10 S NKD-131 25 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 23.2±0.9 24.9±0.9 19.7±1.3 23.8±1.0 23.2±0.8 26.3±0.6 21.2±1.3 22.7±1.3 29.3±1.1 | 0.87±0.15 0.49±0.10 0.59±0.13 0.15±0.12 0.17±0.13 0.37±0.11 |
| NKD-130 10 10 S NKD-134 0 5 S NKD-135 50 10 S NKD-137 50 15 S NKD-141 10 10 S NKD-143 10 10 S NKD-143 10 10 S NKD-145 10 1 S NKD-156 25 10 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 S NKD-163 25 10 S NKD-163 25 10 S NKD-131 25 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 24.9±0.9 19.7±1.3 23.8±1.0 23.2±0.8 26.3±0.6 21.2±1.3 22.7±1.3 29.3±1.1 | 0.49±0.10 0.59±0.13 0.15±0.12 0.17±0.13 0.37±0.11 |
| NKD-134 0 5 S NKD-135 50 10 S NKD-137 50 15 S NKD-141 10 10 S NKD-143 10 10 S NKD-143 10 10 S NKD-145 10 1 S NKD-156 25 10 S NKD-157 10 1 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 PR NKD-163 25 10 S NKD-131 25 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 19.7±1.3 23.8±1.0 23.2±0.8 26.3±0.6 21.2±1.3 22.7±1.3 29.3±1.1 | 0.59±0.13 0.15±0.12 0.17±0.13 0.37±0.11 |
| NKD-135 50 10 S NKD-137 50 15 S NKD-141 10 10 S NKD-143 10 10 S NKD-145 10 1 S NKD-156 25 10 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 PR NKD-163 25 10 S NKD-163 25 10 S NKD-151 25 10 S NKD-153 10 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 23.8±1.0 23.2±0.8 26.3±0.6 21.2±1.3 22.7±1.3 29.3±1.1 | 0.15±0.12 0.17±0.13 0.37±0.11 |
| NKD-137 50 15 S NKD-141 10 10 S NKD-143 10 10 S NKD-145 10 1 S NKD-156 25 10 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 PR NKD-163 25 10 S NKD-163 25 10 S NKD-153 10 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 23.2±0.8 26.3±0.6 21.2±1.3 22.7±1.3 29.3±1.1 | 0.17±0.13 0.37±0.11 |
| NKD-141 10 10 S NKD-143 10 10 S NKD-145 10 1 S NKD-156 25 10 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 PR NKD-163 25 10 S NKD-163 25 10 S NKD-163 25 10 S NKD-151 25 10 S NKD-153 10 10 S NKD-153 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 26.3±0.6 21.2±1.3 22.7±1.3 29.3±1.1 | 0.17±0.13 0.37±0.11 |
| NKD-143 10 10 S NKD-145 10 1 S NKD-156 25 10 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 PR NKD-163 25 10 S NKD-163 25 10 S NKD-163 25 10 S NKD-151 25 10 S NKD-153 10 10 S NKD-153 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 21.2±1.3 22.7±1.3 29.3±1.1 | |
| NKD-145 10 1 S NKD-156 25 10 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 PR NKD-163 25 10 S NKD-163 25 10 S NKD-131 25 10 S NKD-155 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 22.7±1.3 29.3±1.1 | 1.10±0.16 |
| NKD-156 25 10 S NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 PR NKD-163 25 10 S NKD-163 25 10 S NKD-131 25 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 29.3±1.1 | |
| NKD-157 10 1 S NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 PR NKD-151 25 10 S NKD-163 25 10 S NKD-131 25 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | | 0.93±0.17 |
| NKD-159 25 1 S NKD-160 50 10 S NKD-161 25 10 PR NKD-151 25 10 S NKD-163 25 10 S NKD-131 25 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | | 0.63±0.17 |
| NKD-160 50 10 S NKD-161 25 10 PR NKD-151 25 10 S NKD-163 25 10 S NKD-131 25 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 22.6±1.2 | 0.16±0.15 |
| NKD-161 25 10 PR NKD-151 25 10 S NKD-163 25 10 S NKD-131 25 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 26.7±1.1 | 0.25±0.12 |
| NKD-151 25 10 S NKD-163 25 10 S NKD-131 25 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 28.2±0.8 | 0.49±0.12 |
| NKD-163 25 10 S NKD-131 25 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 19.5 ± 1.1 | 0.53 ± 0.11 |
| NKD-131 25 10 S NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 20.8±1.3 | 0.23±0.09 |
| NKD-153 10 10 S NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 23.3±0.7 | 0.77±0.11 |
| NKD-155 10 10 S NKD-133 25 10 PR NKD-144 50 5 S | 26.7±0.8 | 0.50±0.18 |
| NKD-133 25 10 PR NKD-144 50 5 S | 28.4±0.8 | 0.28±0.17 |
| NKD-144 50 5 S | 22.4±1.1 | 1.00±0.16 |
| | 26.8±1.0 | 1.04 ± 0.08 |
| 11112 102 10 10 3 | 20.6±1.1 | 0.42±0.11 |
| NKD-150 0 10 S | 19.0±1.1 | 0.47±0.06 |
| NKD-127 10 10 S | 26.2±0.9 | 0.17±0.14 |
| NKD-129 50 15 S | 24.2±1.1 | 0.28±0.10 |
| NKD-140 10 10 S | 25.3±1.2 | 0.26±0.12 |
| NKD-142 10 5 S | 26.4±1.0 | 0.46±0.11 |
| NKD-148 25 1 S | 27.1±1.1 | 0.34±0.19 |
| NKD-149 25 10 S | 24.1±1.0 | 0.36±0.17 |
| NKD-152 50 10 S | 24.6±1.3 | 0.24±0.15 |
| NKD-158 10 1 S | 25.2±1.3 | 0.28±0.12 |
| NKD-164 25 10 S | 23.3±0.9 | 0.91±0.12 |
| NKD-154 10 10 S | 20.0±1.0 | 1.05 ± 0.08 |
| NK-272 25 5 S | 20.8±0.8 | 0.32±0.09 |
| NK-273 10 5 S | 19.3±0.7 | 0.31±0.12 |
| NK-520 0 10 S | 21.5±0.7 | 0.31±0.15 |
| NK-136 25 10 PR | 20.5±1.1 | 0.23±0.12 |
| NK-129 10 1 PR | 21.8±1.0 | 0.21±0.07 |
| NK-338 25 15 PR | 20.8±1.0 | 0.34±0.15 |
| NK-222 10 15 S | 27.2±1.0 | 0.32±0.11 |
| NK-011 0 15 PR | 19.8±1.3 | 0.55±0.09 |
| NK-292 10 10 S | 20.1±1.2 | 0.49±0.15 |
| NK-201 25 5 S | 20.4±1.0 | 0.56±0.16 |
| NK-515 25 15 S | 23.7±0.8 | 0.30±0.07 |
| NK-069 50 15 S | - | - |
| NK-172 | 22 4 1 1 | 0.26+0.11 |
| LSD (0.05) | 22.4±1.1 | 0.26±0.11 |

Table 2. Resistances, dry matter (DM) and reducing sugar concentrations (RS) in Native Potato Species and old Spanish accessions.

^{*a*} Expressed as percentage of affected tuber area; ^{*b*} S: susceptible PR: partially resistant R: resistant;

In **bold:** accessions resistant or partially resistant

In bold: accessions with dry matter >26% and reducing sugars content <0.20%.

- Not evaluated

* Each value is expressed as the mean \pm standard error (n=3)

susceptible to all assayed pathogens. A total of 5 accessions were resistant and 19 partially resistant against the bacteria S. scabies (≤10% tuber area affected) according to Bouchek-Mechiche et al. (1998). Resistance against S. scabies were found in accessions of the species S. goniocalyx, S. phureja, S. stenotonum, S. tuberosum ssp. andigena, S. tuberosum ssp. tuberosum and the hybrid SxG. For R. solani, 15 accessions belonging to the species S. juzepczukii, S. stenotonum, S. tuberosum ssp. andigena and S. tuberosum ssp. tuberosum were identified as totally or partially resistant ($\leq 5\%$ tuber area affected). Partial resistance to the nematode G. rostochensis (Ro1) was detected in four accessions belonging to Solanum tuberosum ssp. tuberosum and in the NPS accessions NKD-161 (Solanum tuberosum ssp andigena) and NKD-133 (Solanum juzepczukii). The old Spanish accession NK-129 (Solanum tuberosum ssp. tuberosum) was the only accession that showed total or partial resistances against the three tested pathogens.

Table 3 shows the observed variation in DM and RS concentrations. A wide degree of variation in DM was observed, varying between 19.0% and 29.3%. The highest values of DM (>26%) were found in both entries of *S. goniocalix*, in one *S. juzepczukii* in four *S. tuberosum* ssp. andigena accessions (NKD-160, NKD-161, NKD-157 and NKD-143), in three *S. stenotonum* accessions, and in the old Spanish accession NK-222 (*S. tuberosum* ssp. tuberosum). The lowest DM values were detected in most of the accessions belonging to *S. chaucha, S. stenotonum, S. tuberosum* ssp. tuberosum and in the hybrid SxG.

A large variation in reducing sugars (RS) concentration was also observed between, as well as within, *Solanum* species, ranging from 0.19% to 1.10%. The accessions which contained higher RS levels belonged to *S. ajanhuiri* and *S. juzepczukii* species, but also the accession NKD-154 (hybrid between *S. tuberosum* ssp. *andigena* and *S. goniocalyx*) contained high amounts or RS. Low RS

levels were detected in accessions NKD-137, NKD-159 and NKD-157, belonging to *S. tuberosum* ssp. *andigena*. Most of the accessions belonging to *S. stenotonum*, *S. phureja*, *S. goniocalyx*, *S. chaucha* and *S. tuberosum* ssp. *tuberosum* presented medium RS values. However, exceptions could also be found in these species, such as NKD-164 belonging to *S. stenotonum*, and NK-011 and NK-201 belonging to *S. tuberosum* ssp. *tuberosum*.

Differences in the mineral composition are shown in Table 3. Generally, NPS had higher mineral concentrations than the old Spanish accessions. High P concentrations were found in S. goniocalyx, S. ajanhuiri and certain S. tuberosum ssp. andigena accessions (NKD-134, NKD-157, and NKD-160). Accessions of S. goniocalyx showed larger concentrations of macronutrients K, Mg, and Na, and high K values were found in some cultivars of S. tuberosum ssp. andigena. Also, in microelement assays for Fe and Zn, a wide variation was detected in the tubers of these species. Lowest Zn and Fe concentrations were found in 3 S. tuberosum ssp. andigena accessions (NKD-145, NKD-137 and NKD-161). Besides, some accessions with high contents of both elements could be found such as NKD-157, NKD-160 and NKD-134. The highest levels of Zn were observed in NKD-138, whereas the highest values of Fe were found in NKD-157 and NKD-151 belonging to S. chaucha.

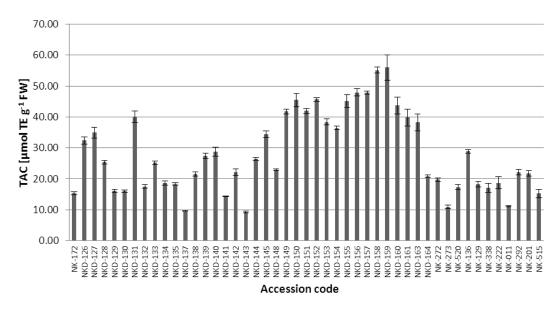
Figure 1 represents the observed variation in TAC and standard errors with a range from 9.29 to 56.00 μ mol TE g⁻¹ FW. In most cases, NPS accessions showed higher antioxidant capacities than the old Spanish ones. In particular, one accession belonging to the species *S. tuberosum* ssp. *andigena* (NKD-159), and another from *S. stenotonum* (NKD-158), revealed the highest antioxidant capacity values.

Figure 2 shows that the total TGA of tubers vary widely between 28.0 and 991.1 mg kg⁻¹ FW. The highest TGA values were found in two *S. tuberosum* ssp. *andigena* accessions (NKD-156 and NKD-157) and in

| Accession code | Р | K | Ca | Mg | Na | Zn | Fe |
|----------------|---------------|---------------|----------------|----------------|-------------------|-----------------|-----------------|
| NKD-138 | 121±1.2 | 668±3.5 | 15.8±0.5 | 33.5±0.6 | 2.32±0.11 | 0.92 ± 0.08 | 1.08 ± 0.10 |
| NKD-139 | 107 ± 1.0 | 574±3.0 | 12.9±0.3 | 36.5±0.8 | 1.60±0.09 | 0.58 ± 0.04 | 0.94 ± 0.09 |
| NKD-126 | 104±0.9 | 499±2.9 | 9.4±0.4 | 28.9±0.6 | 1.87 ± 0.10 | 0.33 ± 0.03 | 0.72 ± 0.08 |
| NKD-128 | 109±1.3 | 636±2.7 | 14.6±0.5 | 23.9±0.7 | 1.97±0.13 | 0.37 ± 0.02 | 0.51±0.03 |
| NKD-130 | 70±1.1 | 314±3.0 | 16.9±0.4 | 14.4±0.7 | 2.30±0.15 | 0.32 ± 0.04 | 0.49 ± 0.04 |
| NKD-134 | 135±1.2 | 801±3.4 | 22.6±0.7 | 42.1±0.9 | 5.43 ±0.18 | 0.73±0.06 | 0.82 ± 0.05 |
| NKD-135 | 90±1.2 | 404±2.8 | 11.9±0.4 | 28.4±0.5 | 3.83±0.13 | 0.30 ± 0.02 | 0.36 ± 0.02 |
| NKD-137 | 61±1.0 | 435±3.0 | 9.1±0.3 | 20.3±0.6 | 2.76±0.10 | 0.25 ± 0.01 | 0.21 ± 0.02 |
| NKD-141 | 84±0.8 | 435±3.1 | 18.9±0.4 | 26.1±0.6 | 3.76±0.14 | 0.43 ± 0.03 | 0.36 ± 0.04 |
| NKD-143 | 93±1.1 | 468±2.7 | 11.9±0.3 | 25.8±0.7 | 3.71±0.14 | 0.53 ± 0.04 | 0.57 ± 0.06 |
| NKD-145 | 66±1.0 | 325±2.9 | 9.1±0.6 | 16.3±0.4 | 1.75±0.09 | 0.25 ± 0.01 | 0.30 ± 0.02 |
| NKD-156 | 68±1.2 | 468±3.2 | 8.9±0.3 | 22.3±0.5 | 2.46±0.11 | 0.30 ± 0.02 | 0.28 ± 0.03 |
| NKD-157 | 140±1.4 | 879±3.4 | 17.0±0.5 | 44.8±0.6 | 3.26±0.12 | 0.86±0.07 | 4.89±0.16 |
| NKD-159 | 103±1.0 | 691±3.0 | 12.2±0.6 | 39.1±0.7 | 2.72±0.11 | 0.65 ± 0.06 | 1.24 ± 0.11 |
| NKD-160 | 110±1.3 | 659±2.7 | 12.8±0.6 | 31.5±0.7 | 2.20 ± 0.08 | 0.78±0.07 | 0.99 ± 0.10 |
| NKD-161 | 108 ± 0.8 | 627±3.2 | 8.1±0.3 | 26.4±0.5 | 3.76±0.12 | 0.22 ± 0.02 | 0.24 ± 0.03 |
| NKD-151 | 104 ± 0.6 | 605±3.0 | 8.6±0.2 | 25.5±0.8 | 1.82 ± 0.08 | 0.59 ± 0.03 | 4.29±0.14 |
| NKD-163 | 74±1.1 | 524±2.8 | 6.3±0.3 | 25.8±0.6 | 1.33 ± 0.07 | 0.21 ± 0.02 | 0.51 ± 0.04 |
| NKD-131 | 116±0.9 | 520±3.1 | 12.5 ± 0.5 | 35.3±0.6 | 2.18 ± 0.06 | 0.56 ± 0.04 | 0.71 ± 0.04 |
| NKD-153 | 121±1.3 | 618±2.9 | 20.1±0.5 | 31.5 ± 0.7 | 4.18 ±0.16 | 0.36 ± 0.03 | 0.45 ± 0.06 |
| NKD-155 | 117 ± 1.2 | 631±2.8 | 21.2±0.6 | 30.5 ± 0.9 | 7.01 ±0.15 | 0.57 ± 0.04 | 0.61 ± 0.05 |
| NKD-133 | 109 ± 1.2 | 513±3.4 | 9.1±0.3 | 29.3±0.4 | 1.77±0.09 | 0.41 ± 0.04 | 0.69 ± 0.06 |
| NKD-144 | 84±0.7 | 440±3.0 | 25.9±0.4 | 25.5±0.5 | 0.97 ± 0.03 | 0.46 ± 0.03 | 2.32 ± 0.11 |
| NKD-132 | 64±0.7 | 424±3.1 | 14.4 ± 0.3 | 17.1±0.3 | 0.47 ± 0.04 | 0.37 ± 0.04 | 1.13 ± 0.09 |
| NKD-150 | 77±0.8 | 493±2.5 | 14.1±0.3 | 25.4±0.5 | 2.71±0.12 | 0.29 ± 0.02 | 0.54 ± 0.06 |
| NKD-127 | 86±1.0 | 518 ± 2.8 | 10.2 ± 0.4 | 27.0±0.6 | 2.06 ± 0.10 | 0.29 ± 0.01 | 0.55 ± 0.07 |
| NKD-129 | 73±1.1 | 460±3.0 | 17.3±0.6 | 18.6±0.4 | 0.93±0.14 | 0.44 ± 0.03 | 1.12 ± 0.10 |
| NKD-140 | 92±1.2 | 476±3.1 | 17.5±0.5 | 21.3±0.5 | 3.63±0.16 | 0.58 ± 0.03 | 0.51 ± 0.06 |
| NKD-142 | 95±1.2 | 531±3.3 | 8.3±0.3 | 25.1±0.5 | 4.36±0.16 | 0.60 ± 0.04 | 0.76 ± 0.04 |
| NKD-148 | 112 ± 1.4 | 536±3.0 | 7.3±0.2 | 35.1±0.7 | 1.77 ± 0.08 | 0.43 ± 0.04 | 0.88 ± 0.07 |
| NKD-149 | 92±1.0 | 514±3.0 | 10.5 ± 0.3 | 26.6±0.5 | 1.99±0.09 | 0.42 ± 0.05 | 0.56 ± 0.05 |
| NKD-152 | 51±1.1 | 360±2.8 | 14.8±0.3 | 13.1±0.3 | 0.98 ± 0.08 | 0.32 ± 0.03 | 0.75 ± 0.06 |
| NKD-158 | 89±1.2 | 545±2.9 | 12.2±0.4 | 27.0±0.4 | 1.90 ± 0.10 | 0.40 ± 0.03 | 1.39±0.09 |
| NKD-164 | 89±0.7 | 536±3.0 | 11.2±0.4 | 31.0±0.5 | 2.18±0.12 | 0.32 ± 0.02 | 0.56 ± 0.06 |
| NKD-154 | 95±1.0 | 499±3.1 | 12.3±0.5 | 28.1±0.5 | 2.69±0.10 | 0.35±0.03 | 1.35 ± 0.11 |
| NK-272 | 58±0.6 | 377±3.1 | 11.2±0.4 | 16.2±0.2 | 0.94 ± 0.05 | 0.35 ± 0.04 | 0.96 ± 0.08 |
| NK-273 | 68±0.7 | 391±2.6 | 7.0±0.2 | 16.1±0.3 | 0.72 ± 0.03 | 0.38 ± 0.04 | 0.69 ± 0.07 |
| NK-520 | 73±0.7 | 398±2.9 | 11.9±0.4 | 17.8±0.2 | 0.32 ± 0.03 | 0.44 ± 0.03 | 1.38 ± 0.12 |
| NK-136 | 50±0.6 | 310±2.8 | 10.6±0.3 | 13.1±0.2 | 0.88 ± 0.06 | 0.33 ± 0.02 | 0.70 ± 0.08 |
| NK-129 | 53±0.6 | 366±2.7 | 18.6±0.6 | 16.5±0.3 | 1.05 ± 0.04 | 0.42 ± 0.04 | 0.82 ± 0.07 |
| NK-338 | 60±0.8 | 421±3.1 | 14.3±0.3 | 22.2±0.6 | 0.18±0.02 | 0.38 ± 0.04 | 1.10±0.09 |
| NK-222 | 55±1.0 | 274±3.0 | 9.1±0.2 | 17.9 ± 0.4 | 0.62 ± 0.04 | 0.33 ± 0.04 | 0.79±0.07 |
| NK-011 | 53±0.9 | 312±3.0 | 9.9±0.3 | 17.8 ± 0.3 | 0.65 ± 0.04 | 0.38 ± 0.03 | 0.68 ± 0.05 |
| NK-292 | 63±1.0 | 337±2.7 | 15.1±0.5 | 21.5±0.4 | 0.31±0.03 | 0.65 ± 0.06 | 1.65±0.12 |
| NK-201 | 41±0.8 | 260±2.5 | 8.5±0.2 | 12.3±0.3 | 0.30±0.02 | 0.21 ± 0.02 | 0.53±0.04 |
| NK-515 | 65±1.1 | 337±2.8 | 10.6±0.3 | 19.9±0.4 | 1.41±0.06 | 0.35±0.03 | 0.89±0.06 |
| NK-172 | 85±1.2 | 450±3.1 | 11.8±0.4 | 22.3±0.5 | 1.03±0.06 | 0.37±0.01 | 0.77±0.06 |
| LSD (0.05) | 13.93 | 7.98 | 1.76 | 1.29 | 0.30 | 0.31 | 0.42 |

 Table 3. Mineral concentrations in Native Potato Species and old Spanish accessions.

All values are expressed as mg/100 g fresh weight. In **bold:** accessions which have the four highest values for macro and microelements content, * Each value is expressed as the mean \pm standard error (n=3)



R area

Figure 1. Total antioxidant capacity (TAC) in Native Potato Species and old Spanish accessions.

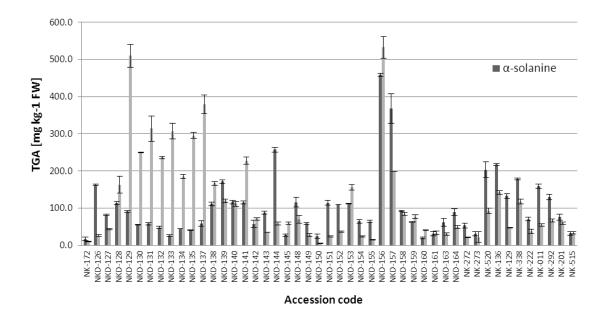


Figure 2. Total glycoalkaloid contents (TGA) in Native Potato Species and old Spanish accessions.

one *S. stenotonum* accession (NKD-129). The lowest TGA levels were observed in the species *S. ajawiri*, *S. phureja*, in the hybrid SxG and in some accessions belonging to *S. tuberosum* ssp. *tuberosum* (`NK-129, NK-338 and NK-201) and *S. stenotonum* (NKD-140, NKD-149 and NKD-164).

The concentration of α -solanine in the 46 cultivars ranged from 16.0 to 459.1 mg kg⁻¹ FW and of α -chaconine from 6.4 to 532.3 mg kg⁻¹ FW (Figure 2). The α -chaconine: α solanine ratio observed in the collection shows a wide variability. Some of the evaluated S. tuberosum ssp. andigena accessions presented high a-chaconine:asolanine ratios (NKD-130, NKD-134 and NKD-135). However, other accessions showed low ratios (NKD-126 and NKD-143), reflecting the large variation in this species. The lowest α -chaconine: α -solanine ratios were found in S. chaucha and S. tuberosum ssp. tuberosum. Some accessions belonging to S. goniocalyx (NKD-155), S. juzepczukii (NKD-144), and the hybrid SxG also showed low α -chaconine: α -solanine ratios.

DISCUSSION

The potato crop is affected by many pests and diseases that cause substantial damages in the field, loss of crops in storage, or affect the quality of the tubers. Managing these pathogens requires crop rotations, utilization of pesticides and other practices that are costly to farmers or environmentally unsafe. However, it may also be possible to confer resistance by transferring a naturally found resistance gene to potato cultivars of interest by conventional hybridization, if possible, or transgenic approaches.

Many complex or partial resistances against *S. scabies*, *R. solani* and *G. rostochensis* were found among the 46 potato accessions of our study. Some old Spanish potatoes also showed very interesting resistances. *S. tuberosum* ssp. *tuberosum* accessions such as NK-129, NK-136, NK-011, NK-273, NK-520 and NK- 292 showed resistances to one or more pathogens. Moreover, the purple fleshed cultivar NK-136 was also resistant to *P. infestans* in leaves (Ritter *et al.*, 2008). The use of these materials instead of *Solanum* wild species could accelerate breeding programmes, due to the absence of interspecific hybridization barriers.

In addition, higher DM values were found among cultivated NPS, particularly in S. tuberosum ssp. andigena accessions. RS concentrations vary widely between, and also within, potato species. In fact, the concentration of glucose and fructose vary depending on variety and storage conditions al.. 2003). (Amrein et These monosaccharides are responsible for the darkening of potato after-cooking and are also implicated in the formation of toxic and carcinogenic acrylamide in the Maillard reaction during frying (Williams, 2005). Mineral analyses have demonstrated that macroelements contents are generally higher in NPS. Moreover, the highest Zn and Fe levels have been found in NPS accessions, but certain old Spanish potatoes also showed high concentrations. Antioxidant capacity assays revealed that NPS had higher TAC values. It is surprising that the highest TAC values were found in yellow or partially purple fleshed accessions, such as NKD-150, NKD-152, NKD-155, NKD-160 and NKD-158, NKD-159 and NKD 160, while none of the purple fleshed accessions showed outstanding values. Brown et al. (2007) also observed that diploid and triploid accessions had larger total carotenoid contents than tetraploids and detected higher lipophilic ORAC values in diploid species. André et al. (2009) observed comparable TAC values in 13 native Andean potato accessions.

Wild potato species tend to have significantly higher glycoalkaloid concentrations (Smith *et al.*, 1996). Also, many cultivated NPS accessions belonging to the species *S. juzepczukii*, *S. curtilobum*, *S. ajawiri*, *S. stenotonum* and *S. tuberosum ssp. andigena* are known to have elevated total glycoalkaloid values. In fact, Andean

farmers have developed a freeze drying process which is commonly used not only for conservation but also for reducing the gycoalkaloid contents of bitter tubers (Burgos et al., 2008). Higher glycoalkaloid contents were mainly observed in accessions belonging to the native species S. tuberosum ssp. andigena. Total glycoalkaloid concentrations found in several NPS and old Spanish accessions were extremely high compared to other studies (Hellenäs, 2001; Kirui et al., 2009; Savage et al., 2000), although it should be noted that our results are based on a single year . In fact, 45.65% of the samples contained more than the recognised safety limit of 200 mg TGA kg⁻¹ FW (OECD, 2002) and four accessions exceeded even 400 mg TGA kg⁻¹ FW. Knuthsen et al. (2009) analyzed 386 potato samples, representative of the Danish market, and only 3 of them contained more than 200 mg TGA kg⁻¹ FW (0.77%).

The principal glycoalkaloids found in potatoes are α -solanine and α -chaconine, comprising about 95% of total glycoalkaloids (Sotelo and Serrano, 2000). In most cases, the contribution of α chaconine to total glycoalkaloid content of tubers was high. This observation coincides with the results obtained by Eltayeb et al. (2003), Friedman (2006) and Tajner-Czopek et al. (2008). However, some old Spanish potato accessions showed relatively high concentrations of α -solanine.

The potential of NPS for generating additional income for local farmers through exploitation commercial has been several recognised and promoted by institutions and organisations. In fact, many potatoes have very suitable native characteristics for potato breeding. The introduction of resistances and quality traits from cultivated native species is easier because wild species are usually difficult to grow in high latitudes and generally do not produce suitable tubers. The amounts of dry matter and minerals and the antioxidant capacity found in certain accessions may contribute to the diffusion of favourable nutritional properties. Some of them have

also an exotic appearance, texture, and flavour which may be particularly attractive to consumers.

Despite the fact that in Europe and USA native potatoes cannot compete with modern S. tuberosum ssp. tuberosum varieties, there is an increasing demand for special and exotic products like Tunta, Tikapapa and also Jalca Chips, which is a snack made from multicoloured native tubers (Devaux et al., 2010). Purple or red coloured potatoes such as Purple Pelisse, Blue Congo, and Purple Majesty are becoming popular in some innovative restaurants and delicatessen markets. Tasty yellow fleshed potatoes derived from S. phureja cultivars are also appreciated. However, some properties of NPS may potentially imply disadvantages for plant breeders and risks for human health. For example, introgressions from NPS can generate potato clones with unfavourable dormancy periods, high glycoalkaloid contents in tubers, much lower vields, late tuberization, small or misshaped and other negative features. tubers, Therefore, potato breeders have to consider the sum of individual characteristics when working with exotic germplasm in order to introgress the desired characteristics from an NPS accession. while minimizing detrimental traits.

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مقاومت به امراض و خاصیت های غذایی سیب زمینی های غده ای بومی و رقم های قدیمی اسپانیایی

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چکیدہ

مقاومت به امراض و آفات بعضی رقم های سیب زمینی مانند سیب زمینی های گونه های بومی از (رشته کوه های) آند (Andes) شناخته شده است. برخی از این نمونه های ثبت شده از لحاظ خاصیت غذایی و پخت و پز مورد توجه هستند. در این پژوهش، به منظور شناخت مقاومت به Streptomyces معنایی و پخت و پز مورد توجه هستند. در این پژوهش، به منظور شناخت مقاومت به *Streptomyces و نیز تعیین تولید ماده* خشک، قندهای احیا شونده، مواد کانی ،غلظت *Globodera rostochiensis و نیز تعیین تولید ماده* خشک، قندهای احیا شونده، مواد کانی ،غلظت *glycoalkaloid و ظرفیت آنتی اکسیدان کل*، مجموعه ای از ۳۵ سیب زمینی گونه های بومی و ۱۱ نمونه ثبت شده از ارقام قدیمی اسپانیایی بررسی شد. نتایج حاکی از تنوع زیاد در میان و در بین گونه های مطالعه شده بود. بیشتر نمونه ها ویژ گی های مطلوبی نشان دادند در حالی که در برخی از آن ها غلظت های *glycoalkaloid* بالا بود. بر پایه این نتایج می توان گفت که بعضی سیب زمینی های گونه های بومی و ارقام قدیمی اسپانیایی به عنوان منبع مقاومت به امراض و ارزش غذایی برای کاربرد در برنامه های اصلاح ژنتیکی استعداد زیادی دارند.